Directory of Services and Tests

DISTRICT OF COLUMBIA
DFS
DEPARTMENT OF FORENSIC SCIENCES

Integrity | Accountability | Trust

401 E Street SW, Washington, DC 20024

Contents

About DC Public Health Laboratory

The District of Columbia (DC) Department of Forensic Science (DFS) Public Health Laboratory Division (PHL) tests samples for a wide variety of matrices from bacterial and viral infections, heavy metals, toxic or volatile materials, and other hazards to public health and safety. The DC PHL provides clinical diagnostic testing, disease surveillance, emergency response support, and other essential services to the District.

Using a multitude of new instrumentation and methods the DC PHL employs chemists, medical technologists, medical technicians, and other specialists to conduct their testing. The PHL staff collaborate with other governmental and professional organizations, not limited to the Centers for Disease Control (CDC), the Association of Public Health Laboratories (APHL), and other federal agencies. In the District, the PHL works closely with the DC Department of Health (DOH), the Office of the Chief Medical Examiner, Department of the Environment and Energy (DOEE) and the Homeland Security and Emergency Management Agency (HSEMA) to protect the District and its residents.

Location and Hours of Operation

Address

Consolidated Forensics Lab
401 E Street SW
Washington, DC 20023

Hours

Monday through Friday: 8:00 a.m. – 5:30 p.m.
Saturday through Sunday: Closed

Contact Information
Daytime Phone: 202- 727-8956

After- hours contact: (202) 868-6561

Email: DFS-PHL-Inquiry@dc.gov

DFS PHL website: https://dfs.dc.gov/page/public-health-laboratory-division-phl

Test Requests

Diagnostic Testing

All diagnostic test requests submitted to the DC PHL must first go through an approval process by the DC Department of Health (DOH). The submitting physician can gain approval by contacting the DOH (EPI on call number, phone call, or doh.epi@dc.gov).

If testing is deemed necessary, the requesting Epidemiologist will inform the DC PHL and the submitter.
Prior to submission of test requests and specimens, requests are to be communicated via email at DFS-PHL-Inquiry@dc.gov or call the laboratory directly at (202) 727-8956.

Please include the following information for initial communication:

- Anticipated number of patients and number of specimens per patient requiring testing.
- Name of facility
- Address of facility for pick-up
- Point of contact for coordinating submission (name, phone #, email)
- Point of contact for results (name, title, phone #, email)
- If needed, Indicated the number of stool collection kits and/or shipping supplies needed for delivery via courier to the facility.
- **Do NOT include patient identifiers in the body of the email.**

Surveillance Testing

The DC PHL performs surveillance testing to monitor the prevalence and spread of diseases or health conditions. Testing is conducted on a regular basis on specimens submitted from area healthcare facilities and community testing sites.

Laboratory Response Network (LRN)

The LRN programs at DC PHL perform testing to screen for potential biothreats, emerging infectious diseases, chemical threats and other public health emergencies.

Test Ordering and Result Reporting

Electronic Test Ordering

LabOnline is an online platform that enables authorized users to place test orders through the DC PHL. LabOnline provides the capability to track sample progression in real-time, as well as securely view, print, and download results and reports.

To register for an account with the DC PHL, please visit the DC PHL's website: https://dfs.dc.gov/publication/phl-forms-and-documents and fill out the LabOnline *User Agreement and Facility Request Form* and submit it to the DC PHL via email: DFS-LabOnline@dc.gov. Once your request has been received and approved by the DC PHL, you will receive an email with your account information and log-in credentials.

LabOnline Sign in page:
https://phl-labonline.dc.gov/Account/SignIn

Manual Test ordering

In settings where electronic test ordering through LabOnline is not available DC PHL also offers manual test ordering. To download manual test requisitions and chains of custody, please visit the DC PHL's website: https://dfs.dc.gov/publication/phl-forms-and-documents and download the corresponding fillable PDF.

Referral Testing

CDC Referral Testing**: test directory** CDC Test Directory
National Jewish Referral Testing (TB) **National Jewish Advanced Diagnostics Laboratory Test Directory**
Other laboratories:

Testing supply requests

DC PHL Courier/Supply Request Form

Supplies provided by DC PHL:

- Respiratory Virus Collection kits
- Category A and B package and shipping supplies

- Candida auris colonization collection kits
- CRE colonization collection kits
- Blood collection kits for suspected Chemical threat
- Mpox collection kits
- Stool collection kits
- Wastewater collection kits
- Potable/Non-potable water collection kits (for Legionella testing)

Specimen Collection and Packaging requirements

Prior to rejecting an unacceptable specimen, the DC PHL will make every effort to communicate with the ordering physician or another healthcare team member before disposing of the specimen. In situations where there are discrepancies on the paperwork, such as a missing date of collection, attempts will be made to reach out to the individual who collected the specimen to obtain a corrected version of the paperwork, which can then be faxed to the laboratory

Specimens will be rejected according to, but not limited to, the following conditions:

- The information on the specimen label does not match the information on the DC PHL test requisition form.
- The specimen has not been transported at the proper temperature.
- The specimen has not been transported in the proper medium.
- The quantity of specimen is insufficient for testing.
- The specimen is leaking.
- The specimen transport tube exceeded the optimum transit time post-collection, and the specimen is not preserved.
- The quality of the specimen is compromised (e.g. sample was damaged, was too old, or was hemolyzed).
- No paperwork is submitted with specimen, or no client information is listed on the paperwork and the client cannot be contacted.
- Evidence suggests the specimen was not collected properly/not collected under the proper conditions.
- Inappropriate specimen sent for test requested.

Packaging

Proper packaging during transport is essential for the preservation of the specimen and the safety of the specimen's handlers. Prior to sending, ensure that the specimen has been collected and packaged under the appropriate storage conditions and in a leak-proof container. For more information on required packaging and transportation temperatures please refer to the table in the transportation section of this document. Additionally, to minimize breakage and cross

contamination, see that glass tubes containing specimens are either surrounded by protective, cushioning materials, or individually contained in biohazard bags.

The DC PHL follows the suggested "Triple Packaging" method for transportation of biological specimens as defined by International Air Transport Association (IATA) and U.S Department of Transportation (DOT) requirements for shipping biological materials.
For additional packaging information, contact DC PHL at (202-727-6929) or Category A and B shipping

Required Documentation

1. The following are **required** documents for all samples submitted to DC PHL:
    1. LabOnline Shipping Manifest or **DC DFS-PHL Test Requisition Form** (link)
    2. External Chain of Custody (COC) Form (link)

2. Some tests require additional documentation; refer to the test menu for requirements.

3. Place **all** paperwork on the outside pocket of the specimen bag or on top of the specimens before closing the lid of the polystyrene foam-insulated, corrugated fiberboard shipper. Ensure that the Chain of Custody can be seen from the outside, but not the Test Requisition Form.

Courier Service

DC PHL employs courier services to pick-up and transfer all biological specimens from local hospitals and clinics. They are DOT/IATA certified to transport Category A and B specimens, handle dry ice, and perform ad hoc deliveries when necessary.
Once specimens are prepared for collection and all necessary documentation has been finalized, the DC PHL provides a scheduled courier service for pick-ups from nine hospital laboratories, Monday - Thursday. Routine pick-up is available at the following locations:

- Children's National Medical Center
- Providence Hospital
- MedStar Washington Hospital Center
- Veterans Affairs Medical Center
- MedStar Georgetown University Hospital
- George Washington University Hospital
- Howard University Hospital
- Sibley Memorial Hospital
- Kaiser Permanente

For all other locations and for ad-hoc requests, complete the following request form: DC PHL Courier Request Form

Courier request form:
https://forms.office.com/g/YvxnK7LbkD

(all fields marked with a * are required)

Diagnostic Test Menu

Serology/Immunology

Measles IgG Immunoassay	
Test Description	The DiaSorin LIAISON® Measles IgG assay uses chemiluminescent assay for the qualitative determination of specific IgG antibodies to measles virus in human serum or plasma samples. It is intended to be used as an aid in the determination of serological status to measles virus.
Analyte(s)	Measles IgG
Acceptable Specimen Type(s)	Serum
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C or -70°C prior to testing and during transport
Methodology	Chemiluminescent Immunoassay
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition

Specimen Labeling	• Specimen must have at least two unique identifiers
	• Specimen should be labeled with the following patient demographics:
	  ○ First and Last Name
	  ○ Date of Birth
	  ○ Collection Date

Transport Medium	Sterile Container, Serum Separator Tube
Rejection Criteria	• Quantity/quality not sufficient for testing • Inappropriate specimen sent for test requested • Specimen demographics do not match the test requisition

	• Grossly hemolyzed or lipemic samples as well as samples containing particulate matter or exhibiting obvious microbial contamination should not be tested and will be rejected
Anticipated Turnaround Time	3 business days
Additional information	Specimen may be forwarded to the Centers for Disease Control and Prevention (CDC) for further testing if necessary or requested.

Microbiology

Aerobic Bacteriology Culture & Identification (OCME only)	
Test Description	Cultivation and identification of bacteria from postmortem specimens received from the Office of the Chief Medical Examiner (OCME).
Analyte(s)	Aerobic bacteria
Acceptable Specimen Type(s)	Post-mortem Blood, Tissue, Cerebral spinal fluid (CSF), Body fluids, and Swabs
Specimen Collection, Storage and Preservation	• CSF- Minimum volume 1ml • Specimens should be stored ambient or at 2-8°C prior to testing and during transport.
Methodology	Conventional culture techniques, MALDI-TOF for identification, Gram Stain
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition
Specimen Labeling	• Specimen must have at least one unique identifier • Specimen should be labeled with the collection date
Transport Medium	• Aerobic and Anaerobic Blood Culture Bottles • Sterile Container
Rejection Criteria:	• Specimen identifiers do not match test request • Any sample rejections must be approved by the Group Manager, Director of Laboratory Operations, or Laboratory Director after consultation with the OCME
Anticipated Turnaround Time	5 business days

Possible Results	Full identification (genus and species) will be performed for significant pathogens, pure culture isolates or a predominating organism in a mixed culture, and at the request of the OCME.
Additional Information	Testing is for the Office of the Chief Medical Examiner (OCME) only.

Biothreat Rule Out and Confirmation (Order Approval Required)	
Test Description	Rule out and confirmatory testing for possible bacterial biothreat agents.
Analyte(s)	• *Bacillus anthracis* • *Brucella species* • *Burkholderia species* • *Yersinia pestis* • *Coxiella burnetii* • *Francisella tularensis* • *Orthopoxvirus* • *Non-variola Orthopoxvirus* • *Viral Hemmoraghic Fever (Ebola, Marburg)*
Acceptable Specimen Type(s)	• Pure bacterial isolate ○ Stored refrigerated or at ambient temperature send to DC PHL as soon as possible and within 7 days of identification • Clinical Specimens - Consult Laboratory for instructions
Specimen Collection, Storage and Preservation	• Isolate - Slant (i.e. TSA slant) or plate media (i.e. blood agar plate) • Clinical Specimens - Consult Laboratory for appropriate collection container and guidelines • Contact the Bioterrorism Unit prior to sample submission.
Methodology	Conventional microbiological methods and Real-time Polymerase Chain Reaction (rt-PCR)
Required Forms	DC PHL External Chain of Custody

	DC PHL Test Requisition Form
Transport Medium	Various; contact the Bioterrorism Unit prior to sample submission.
Anticipated Turnaround Time	Preliminary result (1 Business day) Final report 7- 14 business days
Additional Information	• Isolate/clinical specimen may be forwarded to the Centers for Disease Control and Prevention (CDC) for further testing if necessary • Identification/confirmation of Select Agent(s) will require the submitting lab to complete additional forms to submit to the CDC

Stool Bacterial Culture and Identification - (Order Approval Required, Outbreak ie Only)	
Test Description	Cultivation, isolation, identification of aerobic bacteria from stool specimens submitted for epidemiological investigation and surveillance.
Analyte(s)	*E. coli*, *Salmonella, Vibro, and Shigella*
Acceptable Specimen Type(s)	Raw, unpreserved stool samples
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C prior to testing and during transport
Methodology	Bacterial ID and Culture
Required Forms	DC PHL Surveillance Chain of Custody
Transport Medium	Sterile Container
Additional Rejection Criteria	none
Anticipated Turnaround Time	Varies
Additional Information	Norovirus testing may also be performed by request.

Virology/Molecular

Mpox	
Test Description	Qualitative detection of *Mpox virus* in lesion swabs

Acceptable Specimen Type(s)	Paired dry swabs of body lesions
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C prior to testing and during transport
Methodology	RT-PCR, Xpert Mpox test (FDA EUA)
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition CDC 50.34

Specimen Labeling	• Specimen must have at least two unique identifiers
	• Specimen should be labeled with the following patient demographics:
	  ○ First and Last Name
	  ○ Date of Birth
	  ○ Collection Date
	  ○ Collection Site

Transport Medium	VTM/UTM
Rejection Criteria	• Specimen collection date is greater than 7 days
Anticipated Turnaround Time	3 business days
Additional Information	Presumptive positive samples based on travel to an endemic area will be sent to the Centers for Disease Control and Prevention (CDC) for confirmatory and subtyping.

Measles	
Test Description	Qualitative detection of the Measles virus nucleic acid by Real-Time RT-PCR
Analyte(s)	Measles PCR
Acceptable Specimen Type(s)	Buccal, Nasopharyngeal (NP) or Throat Swabs
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C or -70°C prior to testing and during transport
Methodology	RT-PCR
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition
Transport Medium	Viral Transport Medium
Additional Rejection Criteria	• Quantity not sufficient (≤ 250 µL) for testing

Anticipated Turnaround Time	3 business days
Additional Information	None

Mumps	
Test Description	Qualitative detection and identification of Mumps virus nucleic acid.
Analyte(s)	Mumps PCR
Acceptable Specimen Type(s)	Buccal, Nasopharyngeal (NP) or Throat Swabs
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C or -70°C prior to testing and during transport
Methodology	RT-qPCR
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition
Transport Medium	Viral Transport Medium
Additional Rejection Criteria	• Quantity not sufficient (≤ 250 µL) for testing
Anticipated Turnaround Time	3 business days
Additional Information	None

Norovirus	
Test Description	Qualitative detection of Norovirus nucleic acid by Real-Time RT-PCR.
Analyte(s)	• Norovirus Group 1 • Norovirus Group 2
Acceptable Specimen Type(s)	Raw or unpreserved stool samples
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C prior to testing and during transport
Methodology	RT-PCR, Xpert® Norovirus Assay
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition
Transport Medium	Sterile Container

Additional Rejection Criteria	none
Anticipated Turnaround Time	3 business days
Additional Information	None

SARS-CoV-2	
Test Description	Qualitative detection of SARS-CoV-2 nucleic acids from respiratory specimens.
Analyte(s)	SARS-CoV-2
Acceptable Specimen Type(s)	Nasopharyngeal (NP) Swab, Oropharyngeal (OP) Swab, Nasal Swab (NS)
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C prior to testing and during transport
Methodology	• RT-PCR • Transcription Mediated Amplification (TMA) • Xpert® Multiplex Assay
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition Form
Transport Medium	Viral (VTM) or Universal Transport Medium (UTM)
Additional Rejection Criteria	• Collection date is greater than 72 hours
Anticipated Turnaround Time	3 business days
Additional Information	None

ZIKA Virus	
Test Description	Qualitative detection of RNA from the Zika virus.
Analyte(s)	Zika virus
Acceptable Specimen Type(s)	Urine, Serum, or Plasma
Specimen Collection, Storage and Preservation	2-8°C for up to 7 days
Methodology	Transcription Mediated Amplification (TMA)
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition Form

Transport Medium	Sterile Container, Serum Separator Tube
Additional Rejection Criteria	• Specimen is too old for testing (>7 days) • Quantity not sufficient for testing (<5mL)
Anticipated Turnaround Time	3 business days
Additional Information	None

Toxicology

Urine Drug of Abuse Screening with Reflex to Methadone/EDDP Confirmation	
Test Description	Qualitative 14-drug panel with Creatinine for specimen validity.
Analyte(s)	• 6-AM (6-acetylmorphine) • Amphetamine/ Methamphetamine • Barbiturates • Benzodiazepines • Buprenorphine • Cannabinoids (THC) • Cocaine • Ecstasy (MDMA) • Fentanyl • Methadone • Opiates • Oxycodone • Phencyclidine (PCP) • Tramadol • Creatinine
Acceptable Specimen Type(s)	Urine
Specimen Collection, Storage and Preservation	Ambient up to 48 hours, 2-8°C for up to 7 days
Methodology	Analytical Enzyme Immunoassay (EIA), GC-MS
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition Form
Transport Medium	Sterile Container

Rejection Criteria	• Specimen is too old for testing (>7 days) • Quantity not sufficient for testing (<5mL)
Anticipated Turnaround Time	3 business days
Additional Information	Urine specimens with a Creatinine of less than 20ng/mL are considered dilute and should be resubmitted for testing.

Surveillance and Environmental Test Menu

Microbiology

Food Microbiology Screen (OSSE Only)	
Test Description	Microbiology Screen for total and fecal coliforms in prepared food.
Analyte(s)	Total and Fecal coliform bacteria
Acceptable Specimen Type(s)	Sandwiches, Vegetables, Fruits (not in original manufacturer packaging)
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C or ambient prior to testing and during transport
Methodology	Culture (via Petrifilm) and Identification (MALDI-TOF)
Required Forms	DC PHL Chain of Custody Chain of Custody must indicate where specimens were collected
Transport Medium	Not Applicable
Rejection Criteria	Inappropriate specimen sent for test requested
Additional Information	None

Microbiology Referred Isolates (Surveillance Only)	
Test Description	Cultivation, isolation, identification, susceptibility testing and/or genotyping of referred isolates from clinical laboratories for epidemiological surveillance. Includes isolates for Foodborne (PulseNet), Healthcare Associated Infections (HAI), and other bacterial and fungal isolates of public health concern.

Analyte(s)	Foodborne pathogens (*Salmonella, Shigella, Vibrio, Listeria, Campylobacter*, Shiga-toxin producing *E. coli*, Enteroinvasive *E. coli*) HAI pathogens (Carbapenamase-resistant *Enterobacterales* (CRE), *Acinetobacter baumannii* (CRAB), *Pseudomonas aeruginosa* (CRPA), *Candida auris*)
Acceptable Specimen Type(s)	Bacterial Isolates
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C or ambient prior to testing and during transport
Methodology	Bacterial ID and Culture
Required Forms	DC PHL Surveillance Chain of Custody Submitter Susceptibility Report (if available)
Transport Medium	Bacterial Isolates
Additional Rejection Criteria	None
Additional Information	None

Gonococcal Isolates (Surveillance Only)	
Test Description	Cultivation, isolation and identification of specimens from clinical laboratories for epidemiological surveillance.
Analyte(s)	Neisseria gonorrhoeae
Acceptable Specimen Type(s)	Endocervical, Vaginal, Throat, Rectal and Male Urethral swabs
Specimen Collection, Storage and Preservation	Specimens should be stored ambient prior to testing and during transport
Methodology	Bacterial ID, Gram Stain and Culture
Required Forms	DC PHL Surveillance Test Requisition
Transport Medium	• In-Tray • Gram Stain Slide
Rejection Criteria	• Specimen collection date greater than 72 hours
Additional Information	Samples positive for Neisseria gonorrhoeae may be sent referred for susceptibility testing.

Chlamydia trachomatis and Neisseria gonorrhoeae PCR	
Test Description	The Aptima Combo 2® assay is a target amplification nucleic acid probe test that utilizes target capture for the in vitro qualitative detection and differentiation of ribosomal RNA (rRNA) from Chlamydia trachomatis (CT) and/or Neisseria gonorrhoeae (GC) to aid in the diagnosis of chlamydial and/or gonococcal disease using the Panther® system as specified.
Acceptable Specimen Type(s)	• Endocervical, Vaginal, Throat, Rectal and Male Urethral swabs • Urine
Specimen Collection, Storage and Preservation	Specimens should be stored at 2-8°C prior to testing and during transport
Methodology	Nucleic Acid Amplification (NAAT)
Required Forms	DC PHL Surveillance Test Request
Specimen Labeling	• Specimen must have at least one unique identifier • Specimen should be labeled with the collection date
Transport Medium	• Aptima Unisex Swab Collection Kit • Aptima Urine Specimen Transport Tube
Additional Rejection Criteria	• Collection date greater than 72 hours
Anticipated Turnaround Time	3 business days
Additional Information	Positive samples may be referred for susceptibility testing.

Virology/Molecular

Adenovirus/Human Metapneumovirus, Rhinovirus Panel	
Test Description	The Panther Fusion™ AdV/hMPV/RV assay is a multiplex real-time PCR (RT-PCR) in vitro diagnostic test for the rapid and qualitative detection and differentiation of Adenovirus (AdV), human Metapneumovirus (hMPV), and Rhinovirus (RV). Nucleic acids are isolated and purified from nasopharyngeal (NP) swab specimens obtained from individuals exhibiting signs and symptoms of a respiratory tract infection.
Analyte	Adenovirus/Human Metapneumovirus, Rhinovirus
Acceptable Specimen Type(s)	Nasopharyngeal (NP) Swab
Specimen Collection, Storage and Preservation	Specimens should be stored at 2-8°C prior to testing and during transport

Methodology	RT-PCR
Required Forms	DC PHL External Chain of Custody DC PHL Surveillance Test Requisition

Specimen Labeling	• Specimen must have at least two unique identifiers
	• Specimen should be labeled with the following patient demographics:
	  ○ First and Last Name
	  ○ Date of Birth
	  ○ Collection Date

Transport Medium	• Remel Micro Test M4, M4RT, M5 or M6 Viral Transport Medium • Copan Universal Transport Medium • BD Universal Viral Transport Medium
Rejection Criteria	• Collection date is greater than 96 hours • Inappropriate specimen sent for test requested • Specimen collected in expired transport medium • Specimen identifiers do not match test request
Anticipated Turnaround Time	3 Days
Possible Results	• Detected • Not Detected • Invalid: A new specimen should be collected
Additional Information	None

Influenza A/B/Respiratory Syncytial Virus Panel – Office of the Chief Medical Examiner	
Test Description	The Panther Fusion™ Flu A/B/RSV assay is a multiplex real-time PCR (RT-PCR) in vitro diagnostic test for the rapid and qualitative detection and differentiation of influenza A virus, influenza B virus, and respiratory syncytial virus (RSV). Nucleic acids are isolated and purified from nasopharyngeal (NP) swab specimens obtained from individuals exhibiting signs and symptoms of a respiratory tract infection.
Analyte(s)	Influenza A/B and RSV
Acceptable Specimen Type(s)	Nasopharyngeal (NP) Swab, Oropharyngeal (OP), Postmortem Lung Swabs
Specimen Collection, Storage and Preservation	Specimens should be stored at 2-8°C prior to testing and during transport
Methodology	RT-PCR
Required Forms	DC PHL Chain of Custody

	DC PHL Surveillance Test Requisition
Specimen Labeling	• Specimen must have at least two unique identifiers • Specimen should be labeled with the collection date
Transport Medium	Viral or Universal Transport Medium (VTM/UTM)
Rejection Criteria	• Specimen collection date greater than 96 hours • Specimen collected in expired media • Specimen demographics do not match the test requisition
Additional Information	This test is currently only available for postmortem specimens submitted by the Office of the Chief Medical Examiner (OCME)

Parechovirus/Enterovirus/Adenovirus/RSV Panel	
Test Description	The Panther Fusion™ Parechovirus/Enterovirus/Adenovirus/RSV assay is a multiplex real-time PCR (RT-PCR) in vitro diagnostic test for the rapid and qualitative detection and differentiation of Parechovirus, Enterovirus, Adenovirus and respiratory syncytial virus (RSV). Nucleic acids are isolated and purified from nasopharyngeal (NP) swab specimens obtained from individuals exhibiting signs and symptoms of a respiratory tract infection.
Analyte(s)	Parechovirus, Enterovirus, Adenovirus, RSV
Acceptable Specimen Type(s)	Nasopharyngeal (NP) Swab, Oropharyngeal (OP), Postmortem Lung Swabs
Specimen Collection, Storage and Preservation	Specimens should be stored at 2-8°C prior to testing and during transport
Methodology	RT-PCR
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition
Specimen Labeling	• Specimen must have at least two unique identifiers • Specimen should be labeled with the collection date
Transport Medium	Viral or Universal Transport Medium (VTM/UTM)
Additional Rejection Criteria	• Specimen collection date greater than 96 hours
Additional Information	This test is currently only available for postmortem specimens submitted by the Office of the Chief Medical Examiner (OCME)

Rabies

Test Description	The rabies DFA test qualitatively detects rabies virus in the brain tissue of animals demonstrating rabies behavior. The test involves incubating fluorescent antibodies with rabies suspect brain tissue, allowing the antibodies to bind rabies virus.
Analyte(s)	Rabies virus
Acceptable Specimen Type(s)	Whole Specimen, Whole Brain, Brain Tissue
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C prior to testing and during transport
Methodology	Direct Fluorescent Antibody (DFA)
Required Forms	DC PHL Chain of Custody DC Department of Health Rabies Form
Transport Medium	Sterile Container
Rejection Criteria	Collection date is greater than 72 hours
Anticipated Turnaround Time	1 Business days
Additional Information	None

Respiratory Virus Panel with Reflex to Viral Culture	
Test Description	Qualitative detection and identification of the Influenza A, Influenza B, Respiratory Syncytial Virus (RSV), Adenovirus, Parainfluenza 1, Parainfluenza 2 and Parainfluenza 3 virus in respiratory specimens.
Analyte(s)	• Influenza A • Influenza B • Respiratory Syncytial Virus (RSV) • Adenovirus • Parainfluenza type 1 • Parainfluenza type 2 • Parainfluenza type 3
Acceptable Specimen Type(s)	Nasopharyngeal (NP) Swab, Oropharyngeal (OP) Swab
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C prior to testing and during transport
Methodology	Direct Fluorescent Antibody (DFA) and Viral Culture
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition Form
Specimen Labeling	• Specimen must have at least two unique identifiers

	• Specimen should be labeled with the following patient demographics:
	  ○ First and Last Name
	  ○ Date of Birth
	  ○ Collection Date

Transport Medium	Viral or Universal Transport Medium
Rejection Criteria	• Collection date is greater than 72 hours • Specimen collected on expired media • Information on specimen does not match test request
Anticipated Turnaround Time	3 business days
Possible Results	• Negative • Positive • Invalid: A new specimen should be collected
Additional Information	None

SARS-CoV-2 Wastewater (Environmental Surveillance Only)	
Test Description	Wastewater surveillance is designed to capture the presence of SARS-CoV-2 from raw sewage. The virus is shed by people with and without symptoms allowing for early detection of potential outbreaks.
Analyte(s)	SARS-CoV-2
Acceptable Specimen Type(s)	Raw Wastewater
Specimen Collection, Storage and Preservation	Specimens should be stored at 2–8°C prior to testing and during transport
Methodology	dRT-PCR
Required Forms	DC PHL Chain of Custody
Specimen Labeling	Specimen must have at least one unique identifier
Transport Medium	Sterile Container
Additional Rejection Criteria	• Quantity (>40mL) insufficient for testing
Anticipated Turnaround Time	5 Days
Possible Results	Varies
Additional Information	None

West Nile Virus and Eastern Equine Encephalitis Virus (Surveillance Only)	
Test Description	High throughput RNA extraction and RT-PCR amplification of West Nile virus (WNV) and Eastern Equine Encephalitis virus (EEE) in mosquito pools.
Analyte(s)	WNV and EEE
Acceptable Specimen Type(s)	Mosquito Pool
Specimen Collection, Storage and Preservation	Ambient
Methodology	RT-PCR
Required Forms	DC PHL Chain of Custody
Transport Medium	Sterile Container
Additional Rejection Criteria	None
Anticipated Turnaround Time	5 Days
Additional Information	This test in only available at the request of DCHealth (May-September)

Toxicology

Opioid Surveillance/Biosurveillance	
Test Description	Qualitative and Quantitative opioid
Analyte(s)	• Opioid and Opioid metabolites
Acceptable Specimen Type(s)	Syringe, Urine
Specimen Collection, Storage and Preservation	Ambient up to 48 hours, 2-8°C for up to 7 days
Methodology	GC-MS
Required Forms	DC PHL Chain of Custody
Transport Medium	N/A
Additional Rejection Criteria	N/A
Anticipated Turnaround Time	3 business days

Additional Information	Urine specimens with a Creatinine of less than 20ng/mL are considered dilute and should be resubmitted for testing.

Laboratory Response Network (LRN)

Biothreat

Viral Hemorrhagic Fever Panel (Ebola and Marburg)	
Test Description	Preliminary PCR (Warrior Panel Multiplex RT-PCR) testing for Sudan, Zaire, Reston, Tai Forest, and Bundibugyo ebolaviruses and Marburg viruses, or real time PCR analyses for Zaire ebolavirus, if CDC approves testing; confirmatory testing is conducted at CDC.
Analyte(s)	Ebola and Marburg
Acceptable Specimen Type(s)	Whole Blood, Serum, Plasma
Specimen Collection, Storage and Preservation	Ambient up to 48 hours, 2-8°C for up to 7 days
Methodology	RT-PCR
Required Forms	DC PHL Chain of Custody DC PHL Test Requisition Form
Transport Medium	Sterile Container, EDTA, Serum Separator Tube
Rejection Criteria	• Specimen is too old for testing (>7 days) • Quantity not sufficient for testing (<5mL) • Information on specimen does not match information on test request.
Anticipated Turnaround Time	2 Business Days
Additional Information	This test requires approval prior to submission.

BT Rule Out – Environmental	
Test Description	Cultivation, identification, and confirmation of biological threat agents.
Analyte(s)	• *Bacillus anthracis* • *Brucella* species • *Burkholderia species* • *Yersinia pestis*

	• *Coxiella burnetii* • *Francisella tularensis* • *Bacillus cereus* biovar *anthracis* • *Orthopoxvirus* • Ricin
Acceptable Specimen Type(s)	Various; contact the Bioterrorism Unit prior to sample submission.
Specimen Collection, Storage and Preservation	Various; contact the Bioterrorism Unit prior to sample submission.
Methodology	Bacterial Culture and identification, RT-PCR, immunoassay
Required Forms	DC PHL External Chain of Custody DC PHL Test Requisition Form
Transport Medium	Various; contact the Bioterrorism Unit prior to sample submission.
Additional Rejection Criteria	None
Anticipated Turnaround Time	Varies
Additional Information	This test requires approval prior to submission.